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ENZYMATIC TREATMENT OF PULP TO INCREASE STRENGTH

Background of the Invention

This application is a continuation-in-part of application serial number 09/111,511 entitled Enzymatic Treatment Of Pulp To Increase Strength and filed in the U.S. Patent and Trademark Office on July 8, 1998. The entirety of application serial number 09/111,511 is hereby incorporated by reference.

5 In the manufacture of paper products, such as facial and bath tissues and paper towels, the wet strength and the dry strength of the product are important properties. To achieve these properties, it is common practice to add certain strengthening agents to an aqueous suspension of the papermaking fibers prior to forming the paper sheet. While effective in achieving targeted strength properties, these chemicals are expensive and
10 may be detrimental for other properties (e.g., bulk) or can cause problems for the papermaking process when the whitewater has to be reused.

Therefore, there is a need for a less expensive and more convenient method of improving the sheet strength properties of papermaking fibers.

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Summary of the Invention

It has now been discovered that certain hydrolytic enzymes can randomly react with the cellulose chains at or near the surface of the papermaking fibers to create single aldehyde groups on the fiber surfaces which are part of the fiber. These aldehyde groups, the reducing ends left after random hydrolysis of β -1,4 glucosidic bonds in
20 cellulose, become sites for cross-linking with exposed hydroxyl groups of other fibers when the fibers are formed into sheets and dried, thus increasing sheet strength. In addition, by randomly cutting or hydrolyzing the fiber cellulose chains predominantly at or near the surface of the fiber, degradation of the interior of the fiber cell wall is avoided or at least minimized. Consequently, paper or tissue made from these fibers alone, or made
25 from blends of these fibers with untreated pulp fibers, show an increase in strength properties such as dry tensile, wet tensile, tear, z-direction tensile (surface integrity), etc.

Hence, in one aspect, the invention resides in a method for treating papermaking fibers comprising mixing an aqueous suspension of papermaking fibers and one or more hydrolytic enzymes, optionally in the presence of surfactants, optionally in the presence of
30 other non-cellulolytic enzymes or non-hydrolytic chemical reagents, wherein aldehyde groups are formed predominantly at or near the surface of the fibers.

In another aspect, the invention resides in a method for handling the aqueous suspension of aldehyde-rich, enzyme-treated fibers comprising mechanical beating or kneading if desired, and/or mixing with supplemental chemical additives as needed.

In yet another aspect, the invention resides in a method for making a paper sheet comprising: (a) forming an aqueous suspension of papermaking fibers treated with one or more hydrolytic enzymes capable of randomly hydrolyzing cellulose or hemicellulose to create aldehyde groups; (b) feeding the aqueous suspension into a papermaking headbox; (c) depositing the aqueous suspension onto a forming fabric, whereby the fibers are retained on the surface of the forming fabric in the form of a web while water containing the hydrolytic enzyme(s) passes through the fabric; (d) collecting and recycling the water to recombine the hydrolytic enzyme(s) with additional papermaking fibers to form an aqueous suspension; and (e) drying the web to form a paper sheet.

Particular hydrolytic enzymes useful for purposes of this invention are those enzymes which randomly hydrolyze cellulose and/or hemicellulose to create aldehyde groups. Such enzymes include, without limitation, cellulases, hemicellulases, *endo*-cellulases, *endo*-hemicellulases, carboxymethylcellulases ("CMCases") and *endo*-glucanases. It is known that these enzymes, in particular the cellulases, will degrade the fibrous cell wall, eventually improving pliability, flexibility or softness in coarser webs, but certainly impairing tensile properties at the same time. If these enzymes are not freed of their cellulose binding domain (a step called truncation), they require the presence of a surfactant to moderate the reaction and attain the desired hydrolysis under more controlled conditions. Particularly suitable enzymes for this purpose are truncated *endo*-glucanases and carboxymethylcellulases, which do not require the presence of a surfactant.

For the purposes of this invention, truncated monocomponent *endo*-glucanases or truncated carboxymethylcellulases can be advantageous relative to multi-component cellulases because of their purity (in particular, low or no exocellulase activity) and hence greater treatment control resulting in minimal cell wall damage. However, truncated multicomponent cellulases can also work well, since the reactivity of the *exo*-glucanase portion is severely restricted by chance. A suitable commercially available truncated *endo*-glucanase is sold by Novozymes North America, Inc. (Franklinton, North Carolina), under the name Novozyme® 613, SP 988 or Novozyme® 51016. A related CBD-free CMCase is the commercial preparation EG-40N offered by Clariant Corporation (Charlotte, North Carolina). Still, any other hydrolytic enzymes (natural, modified or even an artificial array of peptides) which possess *endo*-glucanase or carboxymethylcellulase activity can essentially produce similar results.

Suitable papermaking fibers include any virgin or recycled papermaking fibers known in the art, particularly including softwood fibers, such as northern softwood kraft fibers, and hardwood fibers, such as eucalyptus fibers.

As mentioned above, if the hydrolytic enzyme is not truncated, the presence of a
5 surfactant is preferred in the enzyme treatment step for optimal results. A preferred surfactant is a nonionic surfactant, commercially available Tween® 80 (ICI Specialties) or any of the other Tween® 60 series products which are POE sorbitan derivatives. Other suitable nonionic surfactants include DI600® from High Point Chemical Corp.; DI600® is an alkoxyated fatty acid. Furthermore, aryl alkyl polyetheralcohol, e.g. Union Carbide's
10 Triton® X-100 series of surfactants; alkyl phenyl ether of polyethylene glycol, e.g. Union Carbide's Tergitol® series of surfactants; alkylphenoethylene oxide condensation products, e.g. Rhone Poulenc, Incorporated's Igepal® series of surfactants. In some cases an anionic surfactant may be used depending on the type of pulp used. Examples of suitable anionic surfactants are: ammonium or sodium salts of a sulfated ethoxylate
15 derived from a 12 to 14 carbon linear primary alcohol; such as Vista's Alfonic® 1412A or 1412S; and sulfonated naphthalene formaldehyde condensates, e.g. Rohm and Haas's Tamol® SN. In some cases a cationic surfactant can be used, especially when debonding is also desired. Suitable cationic surfactants include imidazole compounds, e.g. Ciba-Geigy's Amasoft® 16-7 and Sapamine® P quaternary ammonium compounds; Quaker
20 Chemicals' Quaker® 2001; and American Cyanamid's Cyanatex®.

The amount of surfactant, if present, can be from about 0.5 to about 6 pounds per metric ton of pulp, more specifically from about 1 to about 5 pounds per metric ton of pulp, more specifically from about 2 to about 4 pounds per metric ton of pulp, and still more specifically from about 2 to about 3 pounds per metric ton of pulp. The specific amount
25 will vary depending upon the particular enzyme being used and the enzyme dosage.

The extent of the hydrolytic modification will depend on the dosage of enzyme applied. The amount of enzyme administered can be denoted in terms of its activity (in enzymatic units) per mass of dry pulp. In general, *endo*-glucanase activity ("CMCase" activity) in cellulases can be assayed by viscosimetry using carboxymethylcellulose (CMC)
30 as a substrate. The higher the activity in a given enzyme preparation, the more pronounced the decay of viscosity will be after a given reaction (incubation) time under predefined experimental conditions. Novo Nordisk Analytical Method 302.1/1-GB, available on request, can be used to assay endoglucanase activity. It calls for the determination of the viscosity loss of a particular solution of CMC (such as Aqualon 7LFD,
35 initial concentration 34g/L) after 30 minutes of incubation with a given enzyme

preparation at pH 7.5 (phosphate buffer) at 40°C. The method relies on the construction of a calibration curve using a standard enzyme of known carboxymethylcellulase activity such as /S, Bagsvaerd Carezyme (batch 17-1196, nominal activity 4931 ECU/g), provided by Novozymes A, Denmark. "ECU" stands for endocellulase units. Determinations of
5 unknown activities are done relative to the standard(s) by interpolation in the calibration curve, with all preparations reacting under the same conditions. The instrument used to measure viscosity reduction is a vibrating rod viscometer, such as the MIVI 6001 unit, manufactured by Sofraser S.A., Villemandeur, France. Still, any other type of viscometer could be used, provided that the same CMC grade is used, a known CMCase standard is
10 employed and the same incubation conditions are followed.

For purposes of this invention, enzyme dosages can vary depending on the desired extent of the treatment and can be from about 5000 to about 200,000 ECU/kilogram of oven dry fibers, more specifically from about 10,000 to about 100,000 ECU/kg, more specifically from about 10,000/kg to about 75,000 ECU/kg, and still more
15 specifically from about 12,000 to about 60,000 ECU/kg. Mixing is desirable to achieve initial homogeneous dispersion and continuous contact between the enzyme and the substrate.

The consistency of the aqueous fiber suspension (weight percent fiber in the total pulp slurry) can be accommodated to meet usual paper mill practices. Low consistencies
20 of about 1% or lower are workable; and consistencies as high as 16% still show sufficient enzyme activity in a pulper. For economical reasons, a consistency in the range of about 8 to about 10% is advantageous.

The reaction conditions for these enzymes can be chosen to provide a pH of about 4 to about 9, more specifically from about 6 to about 8. Temperatures can range from
25 about 0°C (above freezing) to about 70°C. However, it can be envisioned that in the future thermostabilized *endo*-glucanases could react more effectively at extreme temperatures (such as at the boiling point of water), or that alkali-stabilized *endo*-glucanases could react efficiently at high pH ranges (for instance at pH above 11).

Reaction times are also very flexible and depend on the application of enzyme and
30 on the desired extent of the modification. But if kept short, fiber cell wall damage is avoided even with regular cellulases especially in the presence of surfactants. In general, suitable reaction times can be from about 10 to about 180 minutes, more specifically from about 15 to about 60 minutes.

A measure of the effectiveness of the enzyme treatment is the increase in the
35 "copper number" of cellulose. The copper number is defined as the number of grams of copper resulting from the reduction of cupric sulfate by 100 grams of pulp. The procedure

for determining the copper number is described in TAPPI Standard T 430 om-94 "Copper Number of Pulp". Historically, copper number determinations have been used to detect damage to cellulose after hydrolytic or specific oxidative treatments. An increase in reducing groups can indicate deterioration that will have a detrimental impact on

5 mechanical strengths, since the evolution of aldehyde groups has been normally proportional to the random split of the cellulose chain and the decrease of its degree of polymerization throughout the fiber. However, for purposes of this invention, the copper number measures the improvement in the cross-linking ability of the fibers since the chemical modification is substantially restricted to the surface or the surface-near region

10 of the fibers so as to maintain the integrity of the fiber cell walls. In general, the fibers treated in accordance with this invention have a copper number of about 0.10 or more grams of copper per 100 grams of oven-dried pulp, more specifically from about 0.10 to about 1.0 gram of copper per 100 grams of oven-dried pulp, and still more specifically from about 0.15 to about 0.70 gram of copper per 100 grams of oven-dried pulp.

15 The strength increases associated with the treated fibers of this invention, as measured by the dry tensile strength of handsheets made from the treated fibers of this invention compared to the dry tensile strength of handsheets made with untreated fibers, is about 40 percent or greater, more specifically about 50 percent or greater, more specifically about 60 percent or greater, more specifically about 70 percent or greater,

20 more specifically from about 40 to about 150 percent, more specifically from about 50 to about 140 percent, still more specifically from about 60 to about 140 percent, and still more specifically from about 80 to about 140 percent. These strength increases are attributable solely to the enzymatic treatment of the fibers and is without the assistance or contribution of any other supplemental additive(s) or mechanical action that alters the fiber

25 structure, such as refining.

Dried paper made from the treated fibers of this invention can be repulped, a new handsheet formed and dried without significant loss of the dry tensile strength.

Examples

30 Example 1.

In order to illustrate the method of this invention, two different common papermaking fiber pulps were treated with a truncated *endo*-glucanase in accordance with this invention. More specifically, northern softwood bleached kraft fibers, and in a separate experiment, Brazilian eucalyptus bleached kraft pulp fibers, were treated with

35 83,000 ECU/Kg of Novozyme 613® for 15 to 60 minutes in a hydropulper at 8% consistency, 40°C and a pH of 7. The reaction was terminated with the addition of sodium

hypochlorite to deactivate the enzyme. After treatment, the increase of fiber surface aldehyde groups was measured using the copper number determination.

- Table 1 shows the increase of the copper numbers for the two fully bleached kraft pulps before and after treatment of the fibers with Novozyme 613®. The data listed in Table 1 under Reaction Time 0 is an indication for the number of aldehyde groups originally present throughout the fibers and not only for those placed on the fiber surfaces. To avoid the loss in mechanical strength through hydrolysis, it is essential to restrict the extent of chemical modification to the surface of the fibers, so as to maintain the integrity of the cell wall.

10

Table 1
Copper Number Determination After Hydrolysis with Novozyme 613®

Reaction Time (min)	Northern Softwood	Eucalyptus
0	0.06	0.07
30	0.17	0.29
60	0.18	0.32

- As shown by the data, both fiber types underwent an increase in copper number, indicating an increase in the number of aldehyde groups created by the action of the enzyme at the surface or surface-near regions of the fiber.

Example 2.

- In order to illustrate the improvement in strength properties imparted to paper sheets made with the fibers treated in accordance with this invention, handsheets were made from northern softwood bleached kraft pulp and eucalyptus bleached kraft pulp fibers treated with the enzyme as described above (dosage 83,000 ECU/kg of oven-dried fibers). More specifically, handsheets having a basis weight of 60 grams per square meter were prepared by diluting a fiber sample in water to a consistency of 1.2 weight percent in a British Pulp Disintegrator and allowing the dispersed sample to soak for 5 minutes. The sample was then pulped for 5 minutes at ambient temperature, diluted to 0.3 percent consistency and formed into a handsheet on a square (9x9 inches) Valley Handsheet Mold (Voith Inc., Appleton, WI). The handsheet is couched off the mold by

hand using a blotter and pressed wire-side up at 100 pounds per square inch for 1 minute. Then the handsheet was dried wire-side up for 2 minutes to absolute dryness using a Valley Steam Hotplate (Voith Inc., Appleton, WI) and a standard weighted canvas cover having a lead-filled (4.75 pounds) brass tube at one end to maintain uniform tension. The
 5 resulting handsheet was then conditioned in a humidity-controlled room (23°C, 50% relative humidity) prior to testing.

For comparison, the same northern softwood bleached Kraft fibers were treated with 83,000 ECU/Kg of Novozyme 476® –a “full” monocomponent endoglucanase, a CMCase that contains its cellulose binding domain– under identical experimental
 10 conditions.

Testing of the handsheet strength properties involved three different measures: dry tensile strength, wet tensile strength, and tear index.

Dry tensile strength is the peak load measured at the point of failure of a handsheet strip 1 inch wide and 5 inches long in an Instron Testing Machine Mini 55,
 15 running at a loading rate of 0.5 inch per minute.

Wet tensile strength is the peak load measured at the point of failure of a handsheet strip 1 inch wide and 5 inches long in an Instron Testing Machine Mini 55, running at a loading rate of 0.5 inch per minute, where the handsheet strip is wetted thoroughly as described in Tappi Standard T456 om-87.

20 Tear index is measured as described in Tappi Standard T220 sp-96.

Tables 2 and 3 below summarize the results.

Table 2
Northern Softwood Bleached Kraft Pulp Treated with CBD-Free
Endoglucanase

25

Reaction Time	Incremental Dry Tensile Strength Change	Incremental Wet Tensile Strength Change	Incremental Tear Index Change
(min)	%	%	%
0	0	0	0
15	17	-1	44
30	58	33	50
60	66	28	29

Table 3Eucalyptus Bleached Kraft Pulp Treated with CBD-Free Endoglucanase

5

Reaction Time	Incremental Dry Tensile Strength Change	Incremental Wet Tensile Strength Change	Incremental Tear Index Change
(min)	%	%	%
0	0	0	0
15	32	29	-7
30	37	48	46
60	39	20	70

The results show an increase in both dry and wet tensile strengths of the handsheets (either softwood or hardwood fibers) with time of treatment. Tear strength also increased, in contrast with the marked reduction when a full endoglucanase (containing its cellulose binding domain) is used for treatment under the same conditions (see Table 4). Table 4 summarizes the results of treatment of northern softwood Kraft fibers with Novozyme® 476. In this case, tear strength drops dramatically, showing that the intrinsic strength of the fibers has been debilitated. These results are a clear demonstration of the ability of CBD-free endoglucanases to restrict the hydrolytic effect to the outer layers of the fiber, without damage to the bulk phase.

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Table 4Northern Softwood Bleached Kraft Pulp Treated with Full Endoglucanase

Reaction Time	Incremental Tear Index Change
(min)	%
0	0
15	-69
30	-78
60	-83

20

Example 3.

5 In order to further illustrate the improvement in strength properties imparted to paper sheets made with the fibers treated in accordance with this invention, handsheets were made from northern softwood bleached kraft pulp treated with CBD-free endoglucanase Novozyme 988® under experimental conditions as described above (dosage 14,000 ECU/kg of oven-dried fibers). Table 5 below summarizes the results.

10

Table 5Northern Softwood Bleached Kraft Pulp Treated with Novozyme 988®

Reaction Time (min)	Incremental Dry Tensile Strength Change %
0	0.0
30	79
60	111
120	136

15

Example 4.

At the end of the fiber treatment reaction, enzymatic activity can be slowed down by removal of excess liquor (thickening and dilution) which contains the enzyme. Table 6
20 below shows the activity of an original solution and that of a recovered filtrate and a washing liquor.

More specifically, a northern softwood kraft pulp sample (30 g.o.d.) was treated at 5% consistency with a dose of Novozyme® 613 equivalent to 83,000 ECU/kg. After one hour of gentle mixing at 45°C at pH 7, the pulp slurry was filtered under vacuum to form a
25 fiber mat of approx. 15% consistency. The corresponding filtrate of 400mL had an enzyme activity of 2.42 ECU/mL (1). This represents a total activity of 968 ECU or 39% recovery of the initial enzyme activity.

In a continuation of the previous experiment, the filtered pulp was further washed repeatedly by diluting the filtered fiber mat to 5% consistency and re-thickening it to approx. 15%. The produced washings (taken to a total final volume of 3.5Lts.) still showed an enzyme activity of 0.33 ECU/mL (2), corresponding to a cumulative enzyme recovery of 85% of the theoretical amount when added to the activity in the first filtrate (1+2).

The recovered excess liquor can be recycled back into the enzymatic treatment process leading to significant cost reductions through the partial reuse of the enzyme-containing filtrate. If, however, complete inactivation of the enzyme is needed, different physical (e.g., heat) or chemical (e.g., oxidants such as hypochlorite) quenching alternatives are possible to induce irreversible denaturation of any residual enzyme.

Table 6

Enzymatic Activity Novozyme® 613 Solutions Recovered by Filtration

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Sample	Filtrate ECU/mL	Activity ECU	Recovery %
initial	4.35	2490	-
<u>1</u>	2.42	968	39
<u>2</u>	0.33	1155	46
<u>1 + 2</u>		2123	85

The results of Table 6 show that most of the enzyme activity can be recovered using ordinary dewatering.

It will be appreciated that the foregoing examples, given for purposes of illustration, are not to be construed as limiting the scope of the invention, which is defined by the following claims and all equivalents thereto.